

Review Article

Food-chain selenium and human health: emphasis on intake

Margaret P. Rayman

Nutritional Sciences Division, Faculty of Health and Medical Sciences, University of Surrey, Guildford, Surrey GU2 7XH, UK

(Received 12 September 2007 – Revised 16 November 2007 – Accepted 21 November 2007 – First published online 18 March 2008)

Following the publication of the landmark trial of Clark *et al.* in 1996 that appeared to show that Se could reduce the risk of cancer, awareness of the importance of Se to human health has markedly increased. As a result, there is now much more aggressive marketing of Se supplements and functional foods, even in situations where additional consumption of Se is inappropriate. The present review addresses how Se gets into the food chain, the wide variability in Se content of foods and the very different levels of intake between countries and regions. Though it is clear that there are adverse consequences for health of both deficient and excessive intake, health effects at intermediate levels of intake are less certain. Thus it is difficult to define optimal intake which depends on a large number of factors, such as which functions of Se are most relevant to a particular disease state, which species of Se is most prominent in the Se source, which health condition is being considered, the adequacy or otherwise of intake of other nutrients, the presence of additional stressors, and lastly whether the ability to make selenoproteins may be compromised. These complexities need to be understood, particularly by policy makers, in order to make informed judgments. Potential solutions for increasing Se intake, where required, include agronomic biofortification and genetic biofortification or, for individuals, increased intake of naturally Se-rich foods, functional foods or supplements. The difficulties of balancing the risks and benefits in relation to Se intake are highlighted.

Selenium: Intake: Selenium in foods: Selenium and human health: Optimal intake

There is a much greater awareness now of the importance of Se to human health than there was even 10 years ago. This is partly due to the publication of the landmark trial of Clark *et al.*⁽¹⁾ that appeared to show that Se could reduce the risk of cancer. As a result, there is now much more aggressive marketing of Se supplements and functional foods, even in situations where additional consumption of Se is inappropriate.

Both individuals, who take a measure of responsibility for their own health and that of their families, and more importantly, advisory bodies, need to be aware of the complexities surrounding the issue of optimal Se intake in order to make informed judgments. The subject is often treated too simplistically. The present review attempts to air the issues that need to be considered.

Perhaps primarily, individuals need to be aware of the baseline intake in their country or region and whether that intake is adequate or not. There are currently too few data on which to base this judgment, hence companies are able to market Se supplements or functional foods to populations that may already have a perfectly adequate intake of Se. Even in relatively low Se areas, some individuals may consume foods of good Se content (for example, fish) or containing more potent Se species (for example, from garlic, onions or broccoli)

that may give them a higher or more effective intake than might be predicted. An appropriate intake for an individual who is a cigarette smoker or has a family history of prostate cancer may well not be the same as for an individual with a family history of squamous cell carcinoma or diabetes. Individuals may eventually learn whether they have a compromised ability to make selenoproteins, in which case they may need to increase their intake of Se-rich foods.

On the other hand, some evidence is now emerging that links the risk of more subtle adverse health effects to levels of intake well below those known to be toxic. There may even be a possibility of increased risk of one condition even where risk of another is reduced.

An understanding of these niceties requires a certain background knowledge such as: how Se gets into the food chain; the variability of Se content of foods and how that content is affected by food preparation or cooking; how intake varies according to country or region of country; health effects in relation to level of intake and the factors modifying those effects. These issues are addressed below, following which the potential solutions for increasing Se intake, if required, are discussed. Lastly, the difficulties of balancing the risks and benefits in relation to Se intake are highlighted.

Abbreviations: GPx, glutathione peroxidase; NPC, Nutritional Prevention of Cancer; RNI, reference nutrient intake.

Corresponding author: Professor Margaret Rayman, fax +44 1483 300374, email m.rayman@surrey.ac.uk

How selenium gets into the food chain

Se enters the food chain through plants; intake through drinking water is generally trivial⁽²⁾. The amount of Se in foods depends on a number of geological, geographical and other factors. While the Se concentration of the soil is primarily controlled by the underlying geology (carbonatic v. silicatic), the bioavailability of that Se to plants is dependent on pH, redox conditions, amounts of organic matter in the soil, competing ionic species such as sulfate, microbial activity, soil texture, compaction and mineralogy, soil temperature, level of rainfall during the growing season, irrigation and by pedoclimatic variables (temperature and rain intensity excursions) related to fluctuations of soil moisture and pH^(3–10). The uptake of Se by the plant can be greatly inhibited by the simultaneous occurrence of a high soil content of organic matter, Fe hydroxides and clay minerals, all of which can adsorb or bind Se⁽⁴⁾. Se speciation in soils also affects Se bioavailability: selenate is more mobile, soluble and less well adsorbed than selenite⁽⁸⁾. Thus oxidising, alkaline conditions that favour the formation of selenate improve Se bioavailability, while reducing acid conditions that favour the formation of selenite lower bioavailability. According to Fordyce⁽⁸⁾, it is important to understand that even soils that contain adequate or high total Se concentrations can result in Se-deficient crops if the element is not in a form amenable to plant uptake. This is well illustrated by data from the Keshan disease area of Hebei Province, China, that showed a high soil Se content but very low Se bioavailability owing to high organic matter content and lower pH than other soils in the region⁽⁸⁾.

A further important factor is that flowering plant species (angiosperms) differ in their ability to assimilate and accumulate Se. They can be divided into three groups: non-accumulators, Se-indicators (or secondary Se-accumulators) and Se-accumulators⁽¹¹⁾. It appears that the transporters that are responsible for the uptake or translocation of Se are selective such that the ratio of Se:S in the shoots can be higher or lower than that of the solution surrounding the roots⁽¹¹⁾. While non-accumulators rarely accumulate more than 100 µg Se/g dry weight, Se-accumulators can contain up to 40 000 µg Se/g dry weight when grown in Se-rich environments⁽¹¹⁾. The only Se-accumulator plant regularly used as a food source is the tree *Bertholletia excelsa* which produces Brazil nuts, but some crop species of commercial importance can be described as secondary Se-accumulators, for example, *Brassica* species (rapeseed, broccoli, cabbage) and *Allium* species (garlic, onions, leeks and wild leeks)^(11,12). Cereal crops such as wheat, oats, rye and barley are non-accumulators⁽⁸⁾.

The distribution of Se in various parts of the plant depends on species, phase of development and physiological condition⁽¹²⁾. In Se-accumulators, Se accumulates in young leaves during the early vegetative stage of growth but during the reproductive stage it is found at much higher levels in seeds. In non-accumulator cereal crops, there is often about the same amount in grain and roots with smaller amounts in stems and leaves⁽¹²⁾.

Selenium content of foods is very variable

Se concentration in natural food sources has been tabulated by Rayman *et al.*⁽¹³⁾ According to a WHO report⁽¹⁴⁾, the typical

Se content of foods varies as follows: organ meats and seafood, 0.4 to 1.5 µg/g; muscle meats, 0.1 to 0.4 µg/g; most agricultural crops, <1 µg/g dry weight, for example, cereals and grains, less than 0.1 to greater than 0.8 µg/g; dairy products, less than 0.1 to 0.3 µg/g; fruits and vegetables, less than 0.1 µg/g. The variation may be even greater than the above figures imply: for instance in the UK where national sampling of wheat grain has been undertaken over a 16-year period, consistent, extremely low, mean values of 0.025–0.033 ng/g dry weight have been found⁽⁶⁾. Even when grown on seleniferous soils, most vegetables contain a maximum of 6 µg/g and the level in both fruits and vegetables is more likely to be <0.01 µg/g^(15,16).

The variation in Se content of (fresh weight) foods purchased in the upper Midwest of the USA was 72-fold (11–774 µg Se/100 g) for wheat flakes, 57-fold (14–803 µg Se/100 g) for wheat, and 11-fold (19–217 µg Se/100 g) for beef⁽¹⁷⁾ while two brands of the same maize product purchased at the same time from the same store in North America had a 10-fold difference in Se concentration⁽¹⁸⁾. The same foods purchased in different countries may have very different Se content, for example, an average of 57 µg Se/100 g (dry weight) in pasta products made in the USA compared with only 6 µg Se/100 g in Italian pasta⁽¹⁷⁾. Some idea of the Se content of foods purchased in Europe may be obtained by inspecting the values found by Barclay *et al.*⁽¹⁹⁾ who measured the Se content of a range of about 100 foods purchased in the UK between 1993 and 1994. Reilly⁽²⁾ has tabulated Se levels in twelve common foods from a number of countries about the world giving a good illustration of the variability that exists. He also addresses in more detail the Se content of a number of individual foodstuffs: milk, bread and cereals, meat, fish, fruit, vegetables, and Brazil and other nuts.

Brazil nuts are the richest source of food Se, but the content is very variable, ranging from 0.03 to 512 µg/g fresh weight in the studies quoted in the companion paper by Rayman *et al.*⁽¹³⁾ Brazil nuts are harvested from an enormous area of the Amazon basin but soil levels vary from high, in the Menaus to Belem region of the lower Amazon, to low, in the Acre-Rondonia region on the upper Amazon, resulting in high variability in Se content⁽²⁾. Three studies have reported a higher Se content in unshelled than shelled nuts though the reason is not known^(20–22). Two of these studies have drawn attention to the fact that Brazil nuts are exceedingly high in barium, containing levels up to 4000 µg barium/g. Lisk *et al.*⁽²⁰⁾ found that a serving of three Brazil nuts (flesh weight 13.2 g), containing 290 µg Se, also provided 26 mg barium. Barium can be toxic, causing gastroenteritis, muscular paralysis, K deficiency, decreased pulse rate, ventricular fibrillation and extra systoles, and 90% of the barium ingested in that study was retained in the body. The US Environmental Protection Agency's oral reference dose for barium based on toxicological data is 0.2 mg/kg per d, which for a 75 kg individual would be 15 mg/d⁽²⁵⁾. It is clear that this could readily be exceeded by a modest serving of Brazil nuts. Furthermore, Brazil nuts contain small amounts of radium, a radioactive material. Although the amount is very small, typically about 70 (range 3–240) Bq/kg, and most of it is not retained by the body, this is 1000 times higher than in other foods⁽²⁴⁾. Individuals relying on Brazil nuts as their Se source, of

whom there are a not-inconsiderable number, in the UK at least, should be aware both of the uncertainty surrounding the quantity of Se they may be consuming and of the fact that they may be inadvertently consuming barium in amounts exceeding the oral reference dose and radium.

Effect of preparation and cooking on food selenium

According to Fordyce⁽⁸⁾, cooking reduces the Se content of most foods, and studies have shown that vegetables that are normally high in Se such as asparagus and mushrooms can lose 40% during boiling owing to leaching with water. Other studies have estimated that 50% of the Se content is lost from vegetables and dairy products during cooking especially if salt and low-pH components such as vinegar are added, whereas frying foods results in much smaller Se losses^(8,14,25). For Se-enriched *Allium* and *Brassica* plants such as garlic and cabbage, recent studies have estimated that 85 and 89%, respectively, of the total Se is leached into boiling water (H Goenaga Infante, personal communication, 2006). The distribution, concentration and speciation of Se in different edible parts of a plant may well be different: for example, the total Se concentration in the skin of Se-enriched potatoes was found to be almost three times higher than that of the flesh though the highest percentage of Se as selenomethionine (73% of the total Se) was found

in the flesh (H Goenaga Infante, personal communication, 2006). Thus mode of preparation of food must be taken into account when estimating magnitude or nature of Se intake.

Variability in selenium intake by country and region

Intake of Se varies considerably between countries and regions of countries largely owing to the variability of the Se content of plant foods (and hence of animal forage) from one part of the world to another. Se intake data are summarised in Table 1^(7,26–60). Though the level of reliability of such intake data is somewhat variable, it is clear that there is an immense range of intakes, from toxic (approximately = 5 mg/d) in parts of China affected by selenosis (areas of Enshi County, Hubei Province and Ziyang County, Shaanxi Province), through high (Venezuela, parts of North America (North and South Dakota, Montana and Wyoming); approximately = 200–724 µg/d) to high–adequate (rest of North America, Japan; approximately = 100–200 µg/d) to adequate–marginally adequate (Australia, Europe, New Zealand; approximately = 30–90 µg/d) to low or deficient (Eastern European countries, parts of China; approximately = 7–30 µg/d) as judged against current recommendations (tabulated by Rayman⁽⁷⁾). Though plants are the primary source of Se in the diet, animals may be a more reliable source at least for omnivores, as, unlike plants, they

Table 1. Selenium intake data for a number of countries

Country	Se intake (µg/person per d)	Information source
Australia	57–87	Fardy <i>et al.</i> (1989) ⁽²⁸⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Austria	48	Sima & Pfannhauser (1998) ⁽²⁹⁾ (cited by Combs, 2001 ⁽²⁶⁾)
Belgium	28–61	Robberecht & Deelstra (1994) ⁽³⁰⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Brazil	28–37	Maihara <i>et al.</i> (2004) ⁽³¹⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Czech Republic	10–25 (estimate)	Kvíčala <i>et al.</i> (1996) ⁽³²⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Canada	98–224	Gissel-Nielsen (1998) ⁽³³⁾ (cited by Combs, 2001 ⁽²⁶⁾)
China	7–4990	Combs (2001) ⁽²⁶⁾
Croatia	27	Klapec <i>et al.</i> (1998) ⁽³⁴⁾ (cited by Combs, 2001 ⁽²⁶⁾)
Denmark	38–47	Danish Government Food Agency (1995) ⁽³⁵⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Egypt	29	Reilly (1996) ⁽³⁶⁾ (cited by Surai, 2006 ⁽²⁷⁾)
France	29–43	Lamand <i>et al.</i> (1994) ⁽³⁷⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Germany	35	Alfthan & Neve (1996) ⁽³⁸⁾ (cited by Rayman, 2004 ⁽⁷⁾)
India	27–48	Mahalingam <i>et al.</i> (1997) ⁽³⁹⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Ireland	50	Murphy <i>et al.</i> (2002) ⁽⁴⁰⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Italy	43	Allegrini <i>et al.</i> (1985) ⁽⁴¹⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Japan	104–199	Miyazaki <i>et al.</i> (2001) ⁽⁴²⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Nepal	23	Moser <i>et al.</i> (1998) ⁽⁴³⁾ (cited by Surai, 2006 ⁽²⁷⁾)
The Netherlands	39–54	van Dokkum (1995) ⁽⁴⁴⁾ (cited by Rayman, 2004 ⁽⁷⁾)
New Guinea	67	Kumpulainen (1993) ⁽⁴⁵⁾ (cited by Rayman, 2004 ⁽⁷⁾)
New Zealand	20	Donovan <i>et al.</i> (1992) ⁽⁴⁶⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Poland	55–80	Vannoort <i>et al.</i> (2000) ⁽⁴⁷⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Portugal	30–40 (calculated)	Wasowicz <i>et al.</i> (2003) ⁽⁴⁸⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Saudi Arabia	37	Reis <i>et al.</i> (1990) ⁽⁴⁹⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Serbia	15	Al-Saleh <i>et al.</i> (1997) ⁽⁵⁰⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Slovakia	30	Djujic <i>et al.</i> (1995) ⁽⁵¹⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Slovenia	38	Kadrabová <i>et al.</i> (1998) ⁽⁵²⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Spain	30	Pokorn <i>et al.</i> (1998) ⁽⁵³⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Sweden	35	Díaz-Alarcón <i>et al.</i> (1996) ⁽⁵⁴⁾ (cited by Surai, 2006 ⁽²⁷⁾)
Switzerland	31–38	Becker (1989) ⁽⁵⁵⁾ ; Kumpulainen (1993) ⁽⁴⁵⁾ (cited by Rayman, 2004 ⁽⁷⁾)
Turkey	70	Kumpulainen (1993) ⁽⁴⁵⁾ (cited by Rayman, 2004 ⁽⁷⁾)
UK	30–36.5	Reilly (1996) ⁽³⁶⁾ ; Foster & Sumar (1997) ⁽⁵⁶⁾ ; Giray & Hincal (2004) ⁽⁵⁷⁾ (cited by Surai, 2006 ⁽²⁷⁾)
USA	29–39	Ministry of Agriculture, Fisheries and Food (1997) ⁽⁵⁸⁾
Venezuela	106	Food and Nutrition Board (2000) ⁽⁵⁹⁾
	200–350	Combs & Combs (1986) ⁽⁶⁰⁾ (cited by Combs, 2001 ⁽²⁶⁾)

have an absolute requirement for Se which they must get through feed or forage (though it must be remembered that animals, like humans, can be Se deficient⁽⁵⁹⁾). In the UK, for instance, where forage is very low in Se, animal feed is generally supplemented with Se, thereby modestly increasing the Se content of meat and milk. Thus in the UK, meat and poultry make a more important contribution than bread and cereals to dietary Se intake⁽⁶¹⁾. Se is found in highest amounts in organ meats such as kidney and liver while some seafoods contain nearly as much.

Human Se status is dependent not only on the Se content of locally grown foods but also on the extent of use of imported foods. During the 1950s, UK wheat constituted only 15% of the grist⁽¹¹⁾, while wheat imported from Canada, which was much higher in Se content, made a much larger contribution. This situation persisted up to the mid-1980s, but by 2005 the percentage of UK wheat in grists had risen to 80%⁽¹¹⁾. Se intake and status in the UK has fallen in parallel with the decline in imports⁽⁶²⁾ though increased use of sulfur fertilisers (competition of chemically similar species), breeding for higher grain yield per plant, lower atmospheric deposition of Se from coal combustion and the reported decline in cereal consumption are other important factors^(6,11,58). The opposite situation has been seen in New Zealand where Australian wheat with a higher Se content has recently made a significant contribution to Se intake, thereby improving Se status⁽⁶³⁾.

Health effects of selenium in relation to level of intake

Intake of Se ranges from clearly deficient to toxic. At intermediate levels of intake, more subtle health effects have been reported. The situations of deficiency and toxicity are relatively straightforward to describe and will be summarised first. The question of optimal intake for health is much more difficult to address as it requires consideration of the interplay between a large number of factors.

Deficient intake

Overt Se deficiency is associated with Keshan disease, a cardiomyopathy affecting mainly children and women of child-bearing age, frequently fatal, named after the province in the extreme north-east of China where it was endemic⁽⁶⁴⁾. Affected areas had soils that were subject to a strong leaching effect and a high proportion of subsistence farmers who were very dependent on their local food supply^(8,65). The disease occurred in areas where grain crops contained <0.04 µg Se/g and total daily Se intake was between 10 and 15 µg Se/d. Based on epidemiological studies in Keshan disease areas, Chinese workers have suggested a deficiency threshold of 0.02 µg/g in cereal crops for human consumption⁽⁸⁾. Large-scale Se supplementation (0.5–1 mg sodium selenite/week) between 1974 and 1977 dramatically reduced disease incidence⁽⁸⁾. Though the disease was Se-responsive, it is now thought likely that it also had a viral cofactor which in the presence of Se deficiency in the Keshan disease area mutated to a more virulent form that caused the heart condition, as has been shown in the case of Se-deficient mice infected with Cocksackie virus⁽⁶⁶⁾. Cocksackie virus has been isolated from archived heart tissue from patients with Keshan disease⁽⁶⁷⁾.

Though Kashin–Beck disease, an osteoarthropathy found in rural areas of China, Tibet and Siberia, has also been associated with severe Se deficiency, other factors, notably low iodine status, or the presence of fulvic acids or mycotoxins in foods appear likely to be more important^(68,69). More recent data from Tibet appear to support the hypothesis that Kashin–Beck disease occurs as a consequence of oxidative damage to cartilage and bone cells when associated with decreased antioxidant defence, though inhibition of bone remodelling by certain mycotoxins has also been suggested as a potential mechanism⁽⁷⁰⁾.

While levels of Se deficiency of this magnitude are not normally seen in the West, a number of cases of cardiomyopathy, some of which have been shown to be Se-responsive, have been reported in subjects on intravenous nutrition receiving inadequate Se in their infusion solutions^(71,72).

Excessive intake

Overt Se toxicity in humans is far less widespread than Se deficiency⁽⁸⁾. Se toxicity has been studied in animals and observed in humans where signs of selenosis are hair loss, brittle, thickened and stratified nails, garlic breath and skin⁽⁷³⁾. Chronic exposure to high levels of Se has been observed in several populations in seleniferous areas of the world, such as the northern great plains of the USA, parts of Venezuela and Colombia, and one county in China (Enshi, Hubei Province) where the average daily intake of 4.9 mg was associated with a blood Se concentration of 3200 µg/l and symptoms of selenosis. In Enshi, selenosis was associated with the consumption of high-Se crops grown on soils derived from coal containing, on average, greater than 300 µg Se/g (one sample exceeded 80 000 µg/g)⁽⁷⁴⁾. Se from the coal entered the soil by weathering and was available for uptake by crops because of the traditional use of lime as fertiliser in that region. Furthermore, food was cooked and maize was dried over the open flame of this burning coal which also contaminated the atmosphere inside the houses. Morbidity rates reached 50% during peak prevalence years (1961–4) in the worst affected villages which were all located in remote areas among populations of subsistence farmers⁽⁸⁾. The particular outbreak of human selenosis was due to a drought that caused failure of the rice crop, forcing the villagers to eat more high-Se vegetables and maize and fewer protein-rich foods⁽⁷⁴⁾.

Though some plants that grow on seleniferous soils – the Se-accumulators – can take up extremely large amounts of Se ranging from 1000 to 100 000 µg/g (air-dried), farm crops rarely accumulate levels greater than 25–30 µg/g, even in seleniferous areas^(15,16). Based on epidemiological studies in areas affected by selenosis, Chinese workers have suggested a toxicity threshold of 1 µg/g in cereal crops for human consumption⁽⁸⁾. From published data, no health or toxicity problems have been observed up to levels of intake of 819 µg Se/d in China^(75,76) or 724 µg Se/d in the USA⁽⁷⁶⁾. If from cereal or rice, such intake is largely in the form of selenomethionine and selenate⁽⁷⁷⁾. By contrast, the high daily intake of Se in the Inuit of North Greenland (estimated as 193–5885 µg/d), where the diet consists largely of meat and organs from marine mammals, seabirds, fish, and the whales' skin delicacy, muktuk⁽⁷⁸⁾, may include a more substantial

amount of selenocysteine from selenoproteins. Apart from the noted longitudinal striation on the nails, no clinical signs of selenosis have been reported in this population, notwithstanding the extremely high Se intake and blood concentrations well above 1000 $\mu\text{g/l}^{(2)}$: it would appear that Se supplied through a marine diet can be tolerated at levels much higher than normally considered safe. Similarly, despite the Se contamination of the Kesterson National Wildlife Refuge in California and levels of 96 $\mu\text{g/g}$ (wet weight) in fish, up to 130 $\mu\text{g/g}$ (dry weight) in the liver of aquatic birds and up to 5.3 $\mu\text{g/g}$ (wet weight) in the flesh of waterfowl, no adverse health effects were seen in the local population or in domestic animals⁽⁷⁹⁾.

Based on the classic studies of Yang *et al.* in China, the 'low observed adverse effects level' was established as 1540 $\mu\text{g/d}^{(80)}$ and the 'no observed adverse effects level' (NOAEL) as 819 $\mu\text{g/d}^{(75)}$. It should be noted, however, that these values apply only to total Se and may be inaccurate for any specific form. Applying a safety factor to the NOAEL has allowed expert groups in a number of countries to define an upper level of total Se intake believed to be safe. Thus for adults, the 'tolerable upper intake level' for the USA and Canada is 400 $\mu\text{g/d}$, based on a NOAEL of 800 $\mu\text{g/d}^{(59)}$. This same value has been adopted by the WHO⁽²⁵⁾ and is to be adopted by Australia and New Zealand. The 'safe upper limit' in the UK is set at 450 $\mu\text{g/d}$ for adults⁽⁸¹⁾.

Remarkably, in Enshi, China, as described above, Keshan disease and selenosis occur within 20 km of one another; their incidence is dependent on the very different geologies of the two relatively isolated areas⁽⁸⁾.

Optimal intake

Despite food supplies coming from diverse sources, at least in developed countries, there is evidence that in some population groups that Se intake, while not deficient, may be sub-optimal for protection against a number of adverse health conditions. Table 2^(82–123) summarises published studies that showed evidence of an Se-associated health benefit. For each health condition or health effect included in this Table, there is more than one strand of published evidence for a beneficial effect of Se. Where trials are included, they are blinded or double-blinded, randomised and placebo-controlled. Use of data from these studies allows an attempt to be made to estimate optimal intake in relation to specific health benefits.

Ascertaining the optimal intake of Se is not a trivial matter since it is dependent on a number of factors. These include consideration of the mechanism by which Se is thought to act in any particular situation, the species of Se ingested, which type of disease (or which type of cancer) is being considered, the overall nutritional adequacy of the group or population, the extent to which genomic differences between individuals or populations may be relevant, and what other risk or lifestyle factors may be present within the population under consideration. These factors will be considered separately below.

Which function of selenium is being considered?

In the case of the many disease conditions associated with oxidative stress (for example, asthma, rheumatoid arthritis,

pancreatitis, CHD), it would seem important to have an intake of Se that would at least allow full expression of selenoproteins with an antioxidant function. Current recommendations for intake of dietary Se (mean 57 (range 30–85) $\mu\text{g/d}^{(7)}$), hereinafter referred to as the RDA/reference nutrient intake (RDA/RNI), have been set with this objective in mind though we now know that some recommended intakes would be insufficient for the expression of selenoprotein-P, a selenoprotein that appears to have a special role in scavenging peroxynitrite^(124,125).

Furthermore, selenoprotein-P is required for the transport of Se to a number of tissues after its synthesis in the liver⁽¹²⁶⁾ and mouse knock-out studies show its absolute requirement by the brain to avoid neurological dysfunction and brainstem axonal degeneration^(127,128). It would seem, therefore, that Se intake needs to be sufficient to optimise the concentration of plasma selenoprotein-P. Though we do not yet know what level of intake that would require, we do know that current intakes in some parts of Europe, specifically Eastern Europe, and parts of China are inadequate for full expression of glutathione peroxidase (GPx) let alone for full expression of selenoprotein-P⁽¹²⁵⁾.

Apart from the selenoproteins, small-molecular-weight Se compounds such as Se-methyl selenocysteine and γ -glutamyl-Se-methyl selenocysteine are thought to be precursors of the potent anti-cancer agent methyl selenol⁽¹²⁹⁾ which is purported to cause apoptosis, cell-cycle arrest, inhibition of tumour cell invasion and angiogenesis^(130,131). Though small amounts of these compounds are found in members of the *Allium* and *Brassica* families, production of adequate amounts for cancer prevention by metabolism of Se compounds more commonly found in foods probably requires a considerably larger intake, perhaps up to 290 $\mu\text{g Se/d}$, as was the case in Nutritional Prevention of Cancer (NPC) trial subjects⁽¹⁾.

Nature of selenium species in food or supplements consumed

The predominant species of Se in the food (or supplement) consumed will affect the level of intake considered to be optimal, as it will affect bioavailability (absorption and retention), usefulness for synthesis of selenoproteins and ability to produce methyl selenol metabolites. For instance, Se from high-Se broccoli (mainly Se-methyl-selenocysteine, a precursor of methyl selenol) does not accumulate in tissues or increase GPx enzyme activity to the same extent as selenite or selenoamino acids⁽¹³²⁾. Selenite, on the other hand, can be effectively used for selenoprotein synthesis, but it cannot be stored in the body for later use. Selenomethionine (for example, from cereals or high-Se yeast) can act as a storage form of Se in body proteins from which it can slowly be released by catabolism to maintain Se requirements over a longer period. Burk's group has shown⁽¹²⁵⁾ that when Se was supplemented to Chinese subjects in the form of selenomethionine, maximum enzyme activity was reached with a supplement dose of 37 $\mu\text{g/d}$ (on top of a background intake of 10 $\mu\text{g Se/d}$). When the supplement was selenite, a daily dose of 66 μg was required to reach the same maximum level. Thus, Se in the form of selenomethionine was almost twice as effective as Se in the form of selenite in supporting plasma GPx activity. These issues are addressed in depth in the companion paper by Rayman *et al.*⁽¹³⁾

Table 2. Summary of evidence-based health effects of selenium together with an indication of the likely dose-level required blinded, randomised, placebo-controlled trials (RCT), except where specified as double blind

Condition or effect (likely protective intake)	Evidence
Mortality (RDA/RNI)	After adjustment for confounding factors, low plasma Se concentration was significantly associated with higher mortality in the 9-year longitudinal EVA study of 1389 elderly French individuals of mean baseline plasma Se 87 µg/l ⁽⁸²⁾
Cognitive decline (RDA/RNI)	Low serum Se was associated with a significantly higher risk of total mortality in 619 participants in the Women's Health and Aging Study I over a 5-year period (hazard ratio 1.54; 95 % CI 1.03, 2.32) ⁽⁸³⁾ After adjustment for various confounding factors, a significantly increased risk of cognitive decline (OR 1.58; 95 % CI 1.08, 2.31) over a 4-year period was found in French subjects aged 60 to 70 years from the EVA cohort with low plasma Se concentration at baseline ⁽⁸⁴⁾ After controlling for potential confounders, cognitive decline was significantly associated with the magnitude of plasma Se decrease over a 9-year period in the EVA cohort ⁽⁸⁵⁾
Immune system (additional 100–200 µg/d; Europe and USA)	Lower toenail Se was significantly associated with lower cognitive score in rural elderly Chinese ⁽⁸⁶⁾ Supplementation with 100 µg Se/d (as Se-enriched yeast) in an RCT restored the age-related decline in immune response in elderly Belgians ⁽⁸⁷⁾ Supplementation with 200 µg Se/d in an RCT enhanced the cellular immune response of US healthy volunteers and head-and-neck cancer patients ^(88,89) Se supplementation of UK adults with 100 µg Se/d for 15 weeks in a double-blind RCT significantly enhanced the cellular immune response ⁽⁹⁰⁾
Anti-viral effects (additional 100–200 µg/d; China, UK and USA)	Low Se status increases the risk of developing primary liver cancer in hepatitis B/C-positive patients while supplementation of men carrying the hepatitis B surface antigen with 200–500 µg Se/d significantly reduced their risk of developing liver cancer ^(91–94) Se supplementation of UK adults with sodium selenite (50 or 100 µg/d) for 15 weeks in a double-blind RCT resulted in faster clearance of attenuated polio virus with fewer mutations in the viral genome ⁽⁹⁰⁾ In US patients with relatively low Se status (plasma Se < 85 µg/l), HIV infection progressed more rapidly to AIDS with higher mortality ^(95,96) In a double-blind RCT in 187 HIV-positive US adults, 200 µg Se/d caused a marked decrease in hospital admission rates (RR 0.38; <i>P</i> = 0.002) over the 2-year trial ⁽⁹⁷⁾ In a double-blind RCT in 262 HIV-1-seropositive US men and women, the majority of whom were receiving anti-retroviral therapy, Se supplementation (200 µg/d as Se-yeast) significantly suppressed the progression of HIV-1 viral burden and indirectly improved CD4 count ⁽⁹⁸⁾ In a Tanzanian observational study involving 949 HIV-positive pregnant women, mortality decreased by 5 % for every 8 µg/l increase in plasma Se above 85 µg/l (<i>P</i> for trend = 0.01) over a 5-year follow-up period ⁽⁹⁹⁾
Male and female reproduction (RDA/RNI or 100 µg/d)	Sub-fertile Scottish men supplemented with 100 µg Se/d in a double-blind RCT for 3 months had significantly increased sperm motility ⁽¹⁰⁰⁾ Significantly lower serum Se was found in UK women who suffered either first-trimester or recurrent miscarriages compared with women who did not miscarry ^(101,102) UK women in the bottom third of Se status were 4.4 times more likely to develop pre-eclampsia than those in the top two-thirds ⁽¹⁰³⁾
Anti-cancer effects (RDA/RNI or additional 200 µg/d)	Prospective studies have provided evidence for a beneficial effect of Se on risk of lung (meta-analysis ⁽¹⁰⁴⁾), bladder ^(105,106) , oesophageal and gastric cardia cancers ⁽¹⁰⁷⁾ and prostate cancer (for a review, see Rayman ⁽¹⁰⁸⁾ and meta-analyses ^(109,110)) The risk of recurrence of colorectal adenoma, a precancerous condition, in US subjects with baseline serum or plasma Se in the highest quartile (median 150 µg/l), was significantly lower than in those in the lowest quartile (median 113 µg/l) (OR 0.66; 95 % CI 0.50, 0.87) ⁽¹¹¹⁾ Supplementation with 200 µg Se/d in a double-blind RCT (the NPC trial) gave a significant reduction in cancer mortality and in incidence of total cancer, prostate, colorectal and lung cancers ⁽¹⁾ , though in follow-up analyses, only total and prostate cancer incidence remained significant except in the bottom Se tertile ^(112–114) In the above trial, there was a reduced risk of colorectal adenomas in subjects with plasma Se in the bottom tertile (< 105 µg/l) at baseline ⁽¹¹⁵⁾ Chinese RCT with Se as Se-yeast (200 µg/d) or sodium selenite (500 µg/d) have shown that Se supplementation significantly reduces the risk of hepatocellular carcinoma (RR 0.50; 95 % CI 0.35, 0.71) ⁽⁹⁴⁾ There is some evidence that Se may affect not only cancer risk but also progression and metastasis ⁽¹⁰⁸⁾
Protection of the thyroid (additional 200 µg/d; Europe)	In RCT in patients with autoimmune thyroiditis, Se supplementation decreased inflammation and thyroid autoantibody concentrations ^(116–118) An inverse association was found between Se status and thyroid volume, thyroid tissue damage and goitre in French women ⁽¹¹⁹⁾ A positive association was found between the incidence of thyroid cancer and low prediagnostic serum Se concentration in a Norwegian population ^(120,121) In an RCT in 151 women positive for thyroid peroxidase antibodies, supplementation with 200 µg/d Se (as selenomethionine) during pregnancy and the postpartum period reduced thyroid inflammatory activity and the incidence of permanent hypothyroidism ⁽¹²²⁾
CHD (RDA/RNI)	A meta-analysis of twenty-five observational studies showed that a 50 % increase in Se concentrations was associated with a 24 (95 % CI 7, 38) % reduction in CHD risk while in six randomised trials, the pooled RR in a comparison of supplements containing Se with placebo was 0.89 (95 % CI 0.68, 1.17) ⁽¹²³⁾

RNI, reference nutrient intake; EVA, Etude du Vieillissement Artériel; RR, relative risk; NPC, Nutritional Prevention of Cancer.

Which health condition is being considered?

Mortality. As mortality reflects vulnerability to a number of diseases combined, it is worthy of consideration despite the fact that there have been very few studies on plasma Se and mortality in elderly populations. Furthermore, such studies are particularly prone to confounding, as plasma Se concentrations are known to be higher in fit and well-nourished elderly individuals and lower in those who are frail, poorly nourished and unwell⁽¹³³⁾, possibly reflecting a higher level of inflammatory cytokines and lowering of Se in the acute-phase response⁽¹³⁴⁾. Such a criticism cannot be levelled at randomised controlled trials: in a meta-analysis of randomised controlled trials, Bjelakovic *et al.*⁽¹³⁵⁾ found that Se supplementation tended to reduce mortality.

Serum Se was measured at baseline in 619 participants in the Women's Health and Aging Study I (Baltimore, Maryland, 1992–8) and all-cause mortality was determined over a 5-year period. Those with the lowest Se status had a significantly higher risk of total mortality (hazard ratio 1.54, 95% CI 1.03, 2.32)⁽⁸³⁾. Results suggested that the beneficial effect of Se on mortality was linked to a reduction in levels of the inflammatory cytokine, IL-6.

In the 9-year longitudinal Etude du Vieillissement Artériel (EVA) study of 1389 elderly French individuals living independently where various potential confounding factors (socio-demographic characteristics, dietary habits, health, and cognitive factors) were controlled for, low plasma Se concentrations were associated with higher mortality, i.e. for a reduction of 16 µg/l in plasma Se, the relative risk of death was 1.54 (95% CI 1.25, 1.88)⁽⁸²⁾. With a mean plasma Se concentration in the EVA study population of 87 µg/l, a considerable proportion of the participants may not have had a sufficient Se intake for optimal expression of selenoproteins⁽⁶³⁾ including that of selenoprotein-R (methionine sulfoxide reductase), a selenoprotein that has been linked to lifespan⁽¹³⁶⁾. This study therefore suggests that the RDA/RNI level of intake may benefit longevity.

Cognitive function. There can be no doubt that Se is important to the brain⁽¹³⁷⁾: (i) animal models of neurodegenerative disease show enhanced cell loss in Se depletion; (ii) genetic inactivation of cellular GPx increases the sensitivity towards neurotoxins and brain ischaemia while increased GPx activity as a result of increased Se supply or overexpression ameliorates the outcome; (iii) genetic inactivation of selenoprotein-P leads to a marked reduction in brain Se content with a corresponding movement disorder and spontaneous seizures in animal models⁽¹³⁸⁾.

Data from elderly French and Chinese populations of low–moderate Se status (mean baseline plasma Se 86.0 µg/l and mean toenail Se 0.21–0.61 µg/g, respectively) suggest that being at the top rather than the bottom of the low–moderate range of Se status is sufficient to reduce the risk of cognitive decline (see Table 2^(84–86)). This should be achieved by an RDA/RNI level of intake.

However, in the context of cognitive function in the elderly, it should be appreciated that low plasma Se may at least partly reflect a lower production of plasma GPx3 by a less efficient kidney⁽¹³⁹⁾ or lower selenoprotein synthesis resulting from the action of inflammatory cytokines (in the acute-phase response)⁽¹³⁴⁾. Failing kidneys also leak homocysteine, a

known risk factor for dementia, into the bloodstream⁽¹⁴⁰⁾ while in the elderly, hyperhomocysteinaemia is also associated with a pro-inflammatory state⁽¹⁴¹⁾. Whether toenail Se would reflect plasma Se in this context is unknown.

Immune function. The studies in Table 2 show that the cell-mediated immune response can be improved by an additional 100 or 200 µg Se/d even in healthy US volunteers whose baseline Se intake and status are already sufficient to optimise selenoenzyme activity^(87–89). In line with these findings, UK researchers concluded that in the UK population, an additional 100 µg Se/d may be insufficient to support optimal function⁽⁹⁰⁾.

Antiviral effects and HIV. Though animal studies have shown that adequate Se for antioxidant GPx1 activity is important for the avoidance of viral mutation to more virulent forms⁽¹⁴²⁾, the success of supplementation studies with 100 or 200 µg Se/d suggests that this level of intake on top of basic diet may be necessary for antiviral effects in humans^(90,91,93,97,98) (Table 2). It has been suggested that retroviruses such as HIV and Coxsackie B3 have the potential to deplete the host's Se supply by incorporating the Se into viral selenoproteins for their own protection, as has been demonstrated for the DNA virus, *Molluscum contagiosum*^(143–145). Although unproven, this is a potential explanation for the requirement for a Se intake higher than the RDA/RNI.

Fertility and reproduction. The selenoproteins phospholipid GPx4 and sperm nuclei selenoprotein are required for sperm motility and sperm maturation, respectively^(146,147), while selenoprotein-P is required for Se supply to the testes⁽¹⁴⁸⁾. The level of Se intake required to optimise the activities of these selenoproteins is probably somewhere within the range of currently recommended intakes (RDA/RNI), say between 55 and 75 µg/d, as both are high in the hierarchy of selenoprotein expression^(149,150). It follows that the fertility of men whose Se intake is lower than that required to optimise selenoenzyme activity may be improved by supplementation as was demonstrated in sub-fertile Scottish men who showed a significant increase in sperm motility when supplemented with 100 µg Se/d for 3 months⁽¹⁰⁰⁾ (Table 2). There is, however, a suggestion that relatively high intakes (about 300 µg/d) may decrease sperm motility⁽¹⁵¹⁾.

It seems likely that the risk of miscarriage and the pregnancy disease, pre-eclampsia, may also require an intake sufficient to give optimal selenoprotein expression. This can be concluded from studies in UK pregnant women where those with higher Se status had a significantly lower risk of first-trimester or recurrent miscarriage^(101,102) and of pre-eclampsia⁽¹⁰³⁾ (see Table 2). As UK Se intake has been measured as 29–39 µg/d⁽⁵⁸⁾, it is clear that raising it to the RDA/RNI level of intake would be sufficient to optimise female reproductive success.

Cancer. Results of the numerous prospective studies and trials are summarised in Table 2. From prospective studies, the mean or median level of plasma Se required for a significant reduction in cancer risk ranges from > 84 µg/l (for example, for oesophageal and gastric cardia cancer in China⁽¹⁰⁷⁾) to 147 µg/l (for example, for prostate cancer in Hawaii⁽¹⁵²⁾) according to the study, while from trial data, the minimum mean plasma Se for significant reduction in cancer risk in an Eastern US population in the NPC trial ranged from 105 µg/l (all cancers)⁽¹¹²⁾ to 123 µg/l (prostate cancer)⁽¹¹³⁾. The minimum Se

intake required to achieve these plasma concentrations ranges from just below the RDA/RNI level to a total intake of about 140 µg/d from dietary Se (or Se-yeast, which is similarly absorbed and retained⁽¹⁵³⁾). This assertion is based on results of a UK supplementation study in healthy volunteers with a baseline dietary intake of approximately 40 µg/d in which a further 100 µg Se/d as Se-enriched yeast raised plasma Se from 90.3 to 148.4 µg/l⁽¹⁵⁴⁾.

The significant benefit of the Se treatment effect in the NPC trial was restricted to males and to those with baseline plasma Se ≤ 105.2 µg/l. In fact, there was a non-significant increased risk of cancer among those in the highest tertile (baseline plasma Se > 121.6 µg/l) and a significantly increased risk of squamous cell carcinoma in NPC participants with baseline plasma Se in the top two tertiles^(112,155). In addition, further analysis of NPC trial data has shown an increased risk of self-reported type 2 diabetes in those supplemented with Se, though the effect was significant only in those in the top tertile of plasma Se at baseline⁽¹⁵⁶⁾. Though such secondary end-point analyses must be regarded with caution, the advisability of supplementing individuals of already-replete status (say 120–125 µg/l or more⁽¹⁵⁷⁾) with Se must be questioned.

Certainly it should be apparent that in populations that already have a mean baseline intake at the level associated with reduced cancer risk, for example, the Prostate, Lung, Colorectal and Ovarian Cancer Trial population where mean plasma Se was 141.3 µg/l⁽¹⁵⁸⁾, no significant benefit at higher intake or status should be expected, nor indeed was seen in that population. Such populations should not be exposed to additional dietary Se or supplementation.

To date, no cancer trial has used a level of dose that would give a total intake of 140 µg Se/d as suggested above, all having opted for 200 µg Se/d or more.

Thyroid effects. Since the selenoenzymes GPx and thioredoxin reductase are crucial to the protection of the thyroid from the H₂O₂ that is produced there for thyroid hormone synthesis⁽¹⁵⁹⁾ and the selenoenzyme iodothyronine deiodinase is required for the production of active thyroid hormone, it might be expected that an intake of about the RDA/RNI which would optimise the activities of these selenoproteins would be sufficient for the protective effects of Se on the thyroid (see Table 2). However, it was found that sodium selenite or selenomethionine at 200 µg/d was required to decrease inflammation and thyroid autoantibody concentrations in patients with autoimmune thyroiditis, the lower dose of selenomethionine at 100 µg/d being ineffective⁽¹¹⁸⁾. The reason for this rather surprising result is not known.

Coronary heart disease. Evidence that Se affects CHD risk has generally been equivocal despite a good biological rationale for optimal selenoprotein activity and concentration conferring benefit. However, a recent excellent meta-analysis of twenty-five observational studies found the pooled relative risk in a comparison of the highest with the lowest Se concentration categories to be 0.85 (95% CI 0.74, 0.99) in fourteen cohort studies and 0.43 (95% CI 0.29, 0.66) in eleven case-control studies⁽¹²³⁾ though the authors warn that observational studies have provided misleading evidence for other antioxidants. Only two randomised trials have used Se as a single agent, one in Finland that found a significant reduction in risk⁽¹⁶⁰⁾ and the other in the USA that did not⁽¹⁶¹⁾, though

the latter was in an Se-replete population with respect to selenoprotein activity and concentration. Inspection of the serum, plasma and toenail values reported in the meta-analysis together with the randomised trial evidence suggests that achieving the RDA/RNI level of intake may generally be sufficient to reduce CHD risk.

Conclusion on optimal intake for health effects. Partly because of the presence of potential confounding in observational studies from which most of the above data are derived, it is difficult to be categorical about the mean population intake required to minimise the risk of any particular condition (let alone the intake required by any individual when all relevant circumstances, including genotype, are taken into account). However, it does seem clear that the optimal intake of Se depends on the health effect being considered, the risk apparently being reduced at the RDA/RNI level of intake in many cases while others such as cancer and the immune response appear to require a higher intake. Results of many studies are consistent with a threshold effect, i.e. an intake (as represented by serum, plasma or toenail concentration) of Se above which risk is uniformly decreased^(104,107,112,162).

General nutritional adequacy

The intake of other nutrients needs to be taken into account when establishing Se requirements. If a population is well nourished, for instance, with good intake levels of vitamin E and other antioxidant micronutrients, the requirement for Se is likely to be somewhat lower than may be the case for a poorly nourished population such as some of those described in Chinese studies⁽¹⁶³⁾. Thus, the strongest effect of Se on cancer risk has been shown among those subjects with the lowest levels of dietary antioxidant vitamins and carotenoids, and particularly at low α-tocopherol concentrations (for a review, see Rayman⁽¹⁰⁸⁾). Where the population is iodine deficient (for example, the Democratic Republic of Congo), Se intake should not be increased until iodine status has been optimised, as there may be adverse effects on brain development⁽¹⁶⁴⁾.

If ability to make selenoproteins is compromised, additional selenium intake may be needed

The ability to make selenoproteins may be reduced in individuals with failing liver (selenoprotein-P) or kidney (GPx3) function^(139,165). Furthermore, the expression of selenoproteins, particularly of selenoprotein-P, is inhibited by pro-inflammatory cytokines and the acute-phase reaction^(134,166,167). Selenoprotein-P mRNA synthesis is also inhibited by insulin (elevated in certain conditions, for example, obesity) which inactivates the transcription factor FoxO1a that is required for selenoprotein-P promoter activity⁽¹⁶⁸⁾. As selenoprotein-P is largely synthesised in the liver and is the main blood-borne vehicle for transport of Se to other tissues⁽¹³⁶⁾, a reduction in selenoprotein-P expression may have a knock-on effect, reducing the synthesis of selenoproteins in other tissues. By analogy with studies in mice, increased Se intake may be able to compensate for a deficit in selenoprotein activity to some extent^(128,169). With respect to colon cancer risk, the compensation may result from

increased concentration of low-molecular-weight Se metabolites that can produce methyl selenol⁽¹⁶⁹⁾.

Individuals differ substantially in their ability to increase selenoprotein activity in response to additional dietary Se⁽¹⁷⁰⁾. This inter-individual variation may to some extent be accounted for by single nucleotide polymorphisms in selenoprotein genes that determine the efficiency with which individuals can incorporate Se into selenoproteins^(171–174). Selenoprotein synthesis is a complex process requiring multiple factors for the successful insertion of Se as selenocysteine, many of which are encoded by polymorphic genes⁽¹³⁶⁾. This results in inter-individual and inter-racial variation in the efficiency with which selenoproteins are expressed.

A notable example is the *GPx1* gene polymorphism at proline/leucine-198 where possession of the leucine-198 allele is associated with an increased risk of bladder cancer in a Japanese population⁽¹⁷⁵⁾ and of lung cancer in Caucasians but not among ethnic Chinese who do not appear to show this polymorphism⁽¹⁷³⁾. A Danish study found a highly significant correlation between the GPx1 polymorphism and erythrocyte GPx activity such that GPx1 catalytic activity was lowered 5% for each additional copy of the variant leucine-allele ($P=0.0003$)⁽¹⁷⁶⁾. Furthermore, the activity of GPx1 derived from the leucine-containing allele was found to be less responsive to increasing Se supplementation than that from the proline-containing allele⁽¹⁷¹⁾. Thus requirements for dietary Se for optimal protection against cancer may be higher in individuals carrying particular functional selenoprotein single nucleotide polymorphisms (for a review, see Rayman⁽¹⁰⁸⁾).

Epigenetic inactivation of selenoprotein gene expression may also have the potential to alter Se requirements. For instance, a high frequency of GPx3 promoter hypermethylation and progressive loss of GPx3 expression has been found in Barrett's adenocarcinomas and associated lesions⁽¹⁷⁷⁾. GPx3 biallelic hypermethylation and inactivation increased significantly with progression toward neoplasia. It is currently unknown whether increased Se intake can compensate for such loss of selenoprotein expression though the work of Irons *et al.*⁽¹⁶⁹⁾ suggests that that may be the case.

Presence of additional stressors

A number of factors may increase Se requirements and need to be considered when deciding on optimal intake.

Cigarette smokers have higher levels of oxidative stress and lower plasma Se⁽¹³³⁾ and may therefore require a higher Se intake. Similarly, exposure to As, as occurs from drinking water in Bangladesh and Taiwan, may increase the Se requirement since Se can interact with As to reduce its toxicity, possibly by the formation of an Se–As–glutathione conjugate formed in the liver and excreted into bile⁽¹⁷⁸⁾. It is also postulated that a high Hg intake may limit the availability of Se through strong chemical binding⁽⁷²⁾.

Other factors known to be associated with lower Se status that may increase Se requirements are obesity, occurrence of CVD, infection or inflammation^(133,179).

Potential solutions for increasing selenium intake

If further evidence accrues that a certain level of Se intake or status is optimal for reduced disease risk, appropriate solutions

for increasing intake will vary according to whether a public-health or an individual solution is envisaged.

Agronomic fortification with selenised fertilisers was the public-health solution successfully adopted in 1984 by Finland, formerly a low-Se country, that resulted in an increase in Se intake from 38 µg/d (for a daily energy intake of 10 MJ) before fortification to 80 µg/d in 2001^(11,180,181). A similar increase in intake in many European countries would enable populations to achieve recommended Se-intake levels (RDA/RNI). This solution has the merit of using plants as effective buffers, because their growth is reduced at high Se exposure⁽¹⁸²⁾. Other public-health solutions include fortification of the food supply or supplementation of animal feed, which is more effective if the supplement is organic. This last solution again introduces a biological barrier that protects the target population from the effects of accidental overdose⁽¹⁸²⁾.

Genetic biofortification is a more novel solution where food crops are enriched with Se by selecting or breeding crop varieties with enhanced Se-accumulation characteristics⁽¹¹⁾. This method may also minimise the need to use Se fertilisers in all but the lowest soil Se situations. It also has the potential for breeding crop varieties with higher concentrations of specific forms of Se, such as Se-methyl selenocysteine or γ-glutamyl-Se-methyl selenocysteine that can readily be converted to methyl selenol.

Individual solutions may encompass an increased intake of Brazil nuts, offal, fish or shellfish which are good sources of Se⁽²⁾ or the increasingly available and widening range of functional foods that have been created with market demand in mind. High-Se bread, potatoes, garlic, onions, broccoli, beer, tea and mussels can be sourced through the Internet, Se-enriched mushrooms providing a good source of bioavailable Se are now produced in Northern Ireland⁽¹⁸³⁾ while Korea has a chain of restaurants selling pork fed with organic Se ('Selenpork')⁽¹⁸⁴⁾. Se-enriched eggs are produced more than in twenty-five countries worldwide and enjoy a substantial market share in Russia⁽¹⁸⁵⁾. The eggs are claimed to remain fresh for longer⁽¹⁸⁵⁾ while the mushrooms have improved shelf-life⁽¹⁸³⁾.

Supplements are a popular way of increasing Se intake for more affluent consumers. Se from selenomethionine was found to be 1.6 times more bioavailable and much more effective in raising plasma Se than was sodium selenite⁽¹⁵⁷⁾. Se consumed in this way appears to reach its target as shown by significantly increased concentration of Se in prostate tissue in men that consumed Se supplements for up to 1 month^(186–188). Se-methyl selenocysteine is also available as an over-the-counter supplement though there is as yet no published human data on the pharmacokinetics, toxicity or health benefits of this supplement.

According to Taylor & Greenwald⁽¹⁸⁹⁾, at- or near-physiological doses of Se are the appropriate choice in a public-health fortification plan while higher doses might be considered if individual supplementation (or consumption of functional foods) is contemplated.

Striking a balance

Though there are clearly individuals and populations that might benefit from a higher level of intake of Se than they currently

have, the evidence presented in the present review highlights the large number of factors that need to be taken into account before reaching a conclusion on optimal intake. Furthermore, though full knowledge of all the relevant factors in any particular set of circumstances can never be achieved, advisory bodies are obliged to do their best to make appropriate public-health recommendations. An attempt must be made to balance risks and benefits. While there seems no downside to optimising intake to the RDA/RNI level, is it sensible to increase Se intake to the level apparently required to reduce the risk of prostate cancer if it may simultaneously increase the risk of squamous cell carcinoma or type 2 diabetes^(113,155,156)? Potential benefits in terms of immune response, cancer risk and thyroid autoimmune disease must be balanced against the potential risks that may be associated with a supraphysiological intake.

It is very important to be aware of background intake in any particular country or region, as what may be an appropriate additional intake in one country may well be excessive in another. For instance, in those with a background level of intake that already gives them a plasma Se concentration of $\geq 122 \mu\text{g/l}$, cancer risk may potentially be increased with further Se intake⁽¹¹²⁾. This is a concern in the current Selenium and Vitamin E Cancer Prevention Trial (SELECT) where US participants, of mean plasma Se $125 \mu\text{g/l}$ (as found in the Third National Health and Nutrition Examination Survey (NHANES III)⁽¹⁹⁰⁾) are being supplemented with $200 \mu\text{g Se/d}$ as the highly bioavailable selenomethionine⁽¹⁸⁹⁾. Similarly, high users of multivitamin and multimineral supplements should also be aware that they may place themselves at excess risk by topping up their Se intake with additional single supplements⁽¹⁹¹⁾.

With a view to balancing risks and benefits, it would seem sensible to aim just to reach the appropriate threshold level of intake for any particular individual or indeed for a population insofar as it may be judged.

Acknowledgements

Thanks are due to Dr Heidi Goenaga Infante for sharing unpublished results. The author has no conflict of interest to declare.

References

- Clark LC, Combs GF Jr, Turnbull BW, *et al.* (1996) Effects of selenium supplementation for cancer prevention in patients with carcinoma of the skin. A randomized controlled trial. Nutritional Prevention of Cancer Study Group. *JAMA* **276**, 1957–1963.
- Reilly C (2006) *Selenium in Food and Health*, 2nd ed. New York: Springer.
- Diplock AT (1993) Indexes of selenium status in human populations. *Am J Clin Nutr* **57**, Suppl. 2, 256S–258S.
- Fordyce FM, Guangdi Z, Green K & Xiping L (2000) Soil, grain and water chemistry in relation to human selenium-responsive diseases in Enshi District, China. *Appl Geochem* **15**, 117–132.
- Johnson CC, Ge X, Green KA & Liu X (2000) Selenium distribution in the local environment of selected villages of the Keshan Disease belt, Zhangjiakou District, Hebei Province, People's Republic of China. *Appl Geochem* **15**, 385–401.
- Adams ML, Lombi E, Zhao F-J & McGrath S (2002) Evidence of low selenium concentrations in UK bread-making wheat grain. *J Sci Food Agric* **82**, 1160–1165.
- Rayman MP (2004) The use of high-selenium yeast to raise selenium status: how does it measure up? *Br J Nutr* **92**, 557–573.
- Fordyce FM (2005) Selenium deficiency and toxicity in the environment. In *Essentials of Medical Geology*, pp. 373–415 [O Selinus, B Alloway, JA Centeno, RB Finkelman, R Fuge, U Lindh and P Smedley, editors]. London: Elsevier.
- Spadoni M, Voltaggio M, Carcea M, Coni E, Raggi A & Cubadda F (2007) Bioaccessible selenium in Italian agricultural soils: comparison of the biogeochemical approach with a regression model based on geochemical and pedoclimatic variables. *Sci Total Environ* **376**, 160–177.
- Zhao FJ, Lopez-Bellido FJ, Gray CW, Whalley WR, Clark LJ & McGrath SP (2007) Effects of soil compaction and irrigation on the concentrations of selenium and arsenic in wheat grains. *Sci Total Environ* **372**, 433–439.
- Broadley MR, White PJ & Bryson RJ (2006) Biofortification of UK food crops with selenium. *Proc Nutr Soc* **65**, 169–181.
- Terry N, Zayed AM, De Souza MP & Tarun AS (2000) Selenium in higher plants. *Annu Rev Plant Physiol Plant Mol Biol* **51**, 401–432.
- Rayman MP, Goenaga Infante H & Sargent M (2008) Food-chain selenium and human health: spotlight on speciation. *Br J Nutr* **100**, 238–253.
- World Health Organization (1987) *Selenium. A Report of the International Programme on Chemical Safety. Environmental Health Criteria no. 58*. Geneva: WHO.
- Whanger PD (1989) Selenocompounds in plants and their effects on animals. In *Toxicants of Plant Origin, vol. III, Proteins and Amino Acids*, pp. 141–167 [PR Cheeke, editor]. Boca Raton, FL: CRC Press.
- Barceloux DG (1999) Selenium. *J Toxicol Clin Toxicol* **37**, 145–172.
- Keck A & Finley J (2006) Database values do not reflect selenium contents of grain, cereals, and other foods grown or purchased in the upper Midwest of the United States. *Nutr Res* **26**, 17–22.
- Finley JW, Matthys L, Shuler T & Korynta E (1996) Selenium content of foods purchased in North Dakota. *Nutr Res* **16**, 723–728.
- Barclay MNI, MacPherson A & Dixon J (1995) Selenium content of a range of UK foods. *J Food Compos Anal* **8**, 307–318.
- Lisk DJ, Bache CA, Essick LA, Reid CM, Rutzke M & Crown K (1988) Absorption and excretion of selenium and barium in humans from consumption of Brazil nuts. *Nutr Rep Int* **38**, 183–191.
- Chang JC, Gutenmann WH, Reid CM & Lisk DJ (1995) Selenium content of Brazil nuts from two geographic locations in Brazil. *Chemosphere* **30**, 801–802.
- Kannamkumarath SS, Wrobel K, Wrobel K, Vonderheide A & Caruso JA (2002) HPLC-ICP-MS determination of selenium distribution and speciation in different types of nut. *Anal Bioanal Chem* **373**, 454–460.
- United States Environmental Protection Agency (2005) Barium and Compounds, Integrated Risk Information System, CASRN 7440-39-3. <http://www.epa.gov/IRIS/subst/0010.htm>
- Oak Ridge Associated Universities (2007) Brazil Nuts. <http://www.orau.org/PTP/collection/consumer%20products/brazilnuts.htm>
- World Health Organization, Food and Agriculture Organization, International Atomic Energy Agency Expert Group (1996) *Trace Elements in Human Nutrition and Health*. Geneva: WHO.
- Combs GF Jr (2001) Selenium in global food systems. *Br J Nutr* **85**, 517–547.

27. Surai PF (2006) *Selenium in Nutrition and Health*. Nottingham, UK: Nottingham University Press.
28. Fardy JJ, McOrist GD & Farrar YJ (1989) The determination of selenium in the Australian diet using neutron activation analysis. *J Radioanal Nucl Chem* **133**, 391–396.
29. Sima A & Pfannhauser W (1998) Selenium levels in foods produced in Austria. In *Mengen-Spurenelemente, Arbeitstag 18th*, pp. 197–204 [M Anke, editor]. Leipzig: Verlag Harald Schubert.
30. Robberecht HJ & Deelstra HA (1994) Factors influencing blood selenium concentrations: a literature review. *J Trace Elem Electrolytes Health Dis* **8**, 129–143.
31. Maihara VA, Gonzaga IB, Silva VL, Fávoro DIT, Vasconcellos MBA & Cozzolino SMF (2004) Daily dietary selenium intake of selected Brazilian population groups. *J Radioanal Nucl Chem* **259**, 465–468.
32. Kvičala J, Zamrazil V & Jiránek V (1996) Selenium deficient status of the inhabitants of South Moravia. In *Natural Antioxidants and Food Quality in Atherosclerosis and Cancer Prevention*, pp. 177–187 [J Kumpulainen and J Salonen, editors]. Cambridge: Royal Society of Chemistry.
33. Gissel-Nielsen G (1998) Effects of selenium supplementation of field crops. In *Environmental Chemistry of Selenium*, pp. 99–112 [WT Frankenberger Jr and RA Engberg, editors]. New York: Marcel Dekker.
34. Klapac T, Mandić ML, Grigic J, Primorac L, Ikić M, Lovrić T, Grigic Z & Herceg Z (1998) Daily dietary intake of selenium in eastern Croatia. *Sci Total Environ* **217**, 127–136.
35. Danish Governmental Food Agency (1995) *Food Habits of Danes 1995, Main Results*. Søborg, Denmark: Levnedsmiddelstyrelsen.
36. Reilly C (1996) *Selenium in Food and Health*. London: Blackie Academic and Professional.
37. Lamand M, Tressol JC & Bellanger J (1994) The mineral and trace element composition in French food items and intake levels in France. *J Trace Elem Electrolytes Health Dis* **8**, 195–202.
38. Alfthan G & Neve J (1996) Selenium intakes and plasma selenium levels in various populations. In *Natural Antioxidants and Food Quality in Atherosclerosis and Cancer Prevention*, pp. 161–167 [J Kumpulainen and J Salonen, editors]. Cambridge: Royal Society of Chemistry.
39. Mahalingam TR, Vijayalakshni S, Prabhu RK, *et al.* (1997) Studies on some trace and minor elements in blood. A survey of the Kalpakkan (India) population. Part III: Studies on dietary intake and its correlation to blood levels. *Biol Trace Elem Res* **57**, 223–238.
40. Murphy J, Hannon EM, Kiely M, Flynn A & Cashman KD (2002) Selenium intakes in 18–64-y-old Irish adults. *Eur J Clin Nutr* **56**, 402–408.
41. Allegrini M, Lanzola E & Gallorini M (1985) Dietary selenium intake in a coronary heart disease study in Northern Italy. *Nutr Res Suppl.* **1**, 398–402.
42. Miyazaki Y, Koyama H, Hongo T, Sasada Y, Nojiri M & Suzuki S (2001) Nutritional considerations for changes in dietary habit and health promotion practices in community health care; from the viewpoint of selenium (in Japanese with English abstract). *Nippon Kosho Eisei Zasshi* **48**, 243–257.
43. Moser PB, Reynolds RD, Acharya S, Howard MP, Andon MB & Lewis SA (1988) Copper, iron, zinc, and selenium dietary intake and status of Nepalese lactating women and their breast-fed infants. *Am J Clin Nutr* **47**, 729–734.
44. van Dokkum W (1995) The intake of selected minerals and trace elements in European countries. *Nutr Res Rev* **8**, 271–302.
45. Kumpulainen JT (1993) Selenium in foods and diets of selected countries. *J Trace Elem Electrolytes Health Dis* **7**, 107–108.
46. Donovan UM, Gibson RS, Ferguson EL, Ounpuu S & Heywood P (1992) Selenium intakes of children from Malawi and Papua New Guinea consuming plant-based diets. *J Trace Elem Electrolytes Health Dis* **6**, 39–43.
47. Vannoort R, Cressey P & Silvers K (2000) *1997/1998 New Zealand Total Diet Survey. Part 2: Elements*. Wellington, New Zealand: Ministry of Health.
48. Wasowicz W, Gromadzinska J, Rydzynski K & Tomczak J (2003) Selenium status of low-selenium area residents: Polish experience. *Toxicol Lett* **137**, 95–101.
49. Reis MF, Holzbecher J, Martinho E & Chatt A (1990) Determination of selenium in duplicate diets of residents of Pinhel, Portugal, by neutron activation. *Biol Trace Elem Res* **26–27**, 629–635.
50. Al-Saleh I, Al-Doush I & Faris R (1997) Selenium levels in breast milk and cow's milk: a preliminary report from Saudi Arabia. *J Environ Pathol Toxicol Oncol* **16**, 41–46.
51. Djuric I, Djuric B & Trajkovic L (1995) Dietary intake of selenium in Serbia: results for 1991. Naučni Skupovi (Srpska Akademija Nauka i Umetnosti). *Odeljenje Prirodno-Matematičkih Nauka* **6**, 81–87.
52. Kadrová J, Mad'arič A & Ginter E (1998) Determination of the daily selenium intake in Slovakia. *Biol Trace Element Res* **61**, 277–286.
53. Pokorn D, Stibilj V, Gregoric B, Dermelj M & Stupar J (1998) Elemental composition (Ca, Mg, Mn, Cu, Cr, Zn, Se and I) of daily diet samples from some old people's homes in Slovenia. *J Food Compos Anal* **11**, 47–53.
54. Díaz-Alarcón JP, Navarro-Alarcón M, López-García de la Serrana H & López-Martínez MC (1996) Determination of selenium in meat products by hydride generation atomic absorption spectrophotometry – selenium levels in meat, organ meats and sausages in Spain. *J Agric Food Chem* **44**, 1494–1497.
55. Becker W (1989) *Food Habits and Nutrient Intake in Sweden 1989*. Uppsala, Sweden: Swedish National Food Administration.
56. Foster LH & Sumar S (1997) Selenium in health and disease: a review. *Crit Rev Food Sci Nutr* **37**, 211–228.
57. Giray B & Hincal F (2004) Selenium status in Turkey. *J Radioanal Nucl Chem* **259**, 447–451.
58. Ministry of Agriculture, Fisheries and Food (1997) *Ministry of Agriculture, Fisheries and Food, October 1997, Food Surveillance Information Sheet, no. 126. Dietary Intake of Selenium*. London: Joint Food Safety and Standards Group.
59. Food and Nutrition Board & Institute of Medicine (2000) *Dietary Reference Intakes: Vitamin C, Vitamin E, Selenium, and Carotenoids*. Washington, DC: National Academy Press.
60. Combs GF Jr & Combs SB (1986) The biological availability of selenium in foods and feeds. In *The Role of Selenium in Nutrition*, pp. 127–177 New York: Academic Press.
61. Rayman MP & Callahan A (2006) *Nutrition and Arthritis*. Oxford, UK: Blackwell Publications.
62. Rayman MP (1997) Dietary selenium: time to act. *BMJ* **314**, 387–388.
63. Thomson CD (2004) Selenium and iodine intakes and status in New Zealand and Australia. *Br J Nutr* **91**, 661–672.
64. Cheng YY & Qian PC (1990) The effect of selenium-fortified table salt in the prevention of Keshan disease on a population of 1.05 million. *Biomed Environ Sci* **3**, 422–428.
65. Li C (2007) Selenium deficiency and endemic heart failure in China: a case study of biogeochemistry for human health. *Ambio* **36**, 90–93.
66. Levander OA & Beck MA (1997) Interacting nutritional and infectious etiologies of Keshan disease. Insights from Cox-sackie virus B-induced myocarditis in mice deficient in selenium or vitamin E. *Biol Trace Elem Res* **56**, 5–21.

67. Su C, Gong C, Li Q, Chen L, Zhou D & Jin Q (1979) Preliminary results of viral etiology of Keshan disease. *Chin Med J* **59**, 466–472.
68. Moreno-Reynes R, Suetens C, Mathieu F, Begaux F, Zhu D, Rivera MT, Boelaert M, Nève J, Perlmutter N & Vanderpas J (1998) Kashin–Beck osteoarthropathy in rural Tibet in relation to selenium and iodine status. *New Eng J Med* **339**, 1112–1120.
69. Coppinger RJ & Diamond AM (2001) Selenium deficiency and human disease. In *Selenium: Its Molecular Biology and Role in Human Health*, pp. 219–233 [DL Hatfield, editor]. Dordrecht, The Netherlands: Kluwer Academic Publishers.
70. Suetens C, Moreno-Reyes R, Chasseur C, Mathieu F, Begaux F, Haubruge E, Durand MC, Neve J & Vanderpas J (2001) Epidemiological support for a multifactorial aetiology of Kashin–Beck disease in Tibet. *Int Orthop* **25**, 180–187.
71. Huttunen JK (1997) Selenium and cardiovascular diseases – an update. *Biomed Environ Sci* **10**, 220–226.
72. Cooper LT, Rader V & Ralston NV (2007) The roles of selenium and mercury in the pathogenesis of viral cardiomyopathy. *Congest Heart Fail* **13**, 193–199.
73. Whanger P, Vendeland S, Park YC & Xia Y (1996) Metabolism of subtoxic levels of selenium in animals and humans. *Ann Clin Lab Sci* **26**, 99–113.
74. Yang GQ, Wang SZ, Zhou RH & Sun SZ (1983) Endemic selenium intoxication of humans in China. *Am J Clin Nutr* **37**, 872–881.
75. Yang G & Zhou R (1994) Further observations on the human maximum safe dietary selenium intake in a seleniferous area of China. *J Trace Elem Electrolytes Health Dis* **8**, 159–165.
76. Longnecker MP, Taylor PR, Levander OA, *et al.* (1991) Selenium in diet, blood, and toenails in relation to human health in a seleniferous area. *Am J Clin Nutr* **53**, 1288–1294.
77. Whanger PD (2002) Selenocompounds in plants and animals and their biological significance. *J Am Coll Nutr* **21**, 223–232.
78. Hansen JC & Sloth Pederson H (1986) Environmental exposure to heavy metals in N. Greenland. *Arct Med Res* **41**, 21–34.
79. Fan AM, Book SA, Neutra RR & Epstein DM (1988) Selenium and human health implications in California's San Joaquin Valley. *J Toxicol Environ Health* **23**, 539–559.
80. Yang G, Yin S, Zhou R, Gu L, Yan B, Liu Y & Liu Y (1989) Studies of safe maximal daily dietary Se-intake in a seleniferous area in China. Part II: Relation between Se-intake and the manifestation of clinical signs and certain biochemical alterations in blood and urine. *J Trace Elem Electrolytes Health Dis* **3**, 123–130.
81. Expert Group on Vitamins and Minerals (2003) *Safe Upper Levels for Vitamins and Minerals*. London: Food Standards Agency.
82. Akbaraly NT, Arnaud J, Hininger-Favier I, Gourlet V, Roussel AM & Berr C (2005) Selenium and mortality in the elderly: results from the EVA study. *Clin Chem* **51**, 2117–2123.
83. Walston J, Xue Q, Semba RD, Ferrucci L, Cappola AR, Ricks M, Guralnik J & Fried LP (2006) Serum antioxidants, inflammation, and total mortality in older women. *Am J Epidemiol* **163**, 18–26.
84. Berr C, Balansard B, Arnaud J, Roussel AM & Alperovitch A (2000) Cognitive decline is associated with systemic oxidative stress: the EVA study. Etude du Vieillissement Arteriel. *J Am Geriatr Soc* **48**, 1285–1291.
85. Akbaraly NT, Hininger-Favier I, Carriere I, Arnaud J, Gourlet V, Roussel AM & Berr C (2007) Plasma selenium over time and cognitive decline in the elderly. *Epidemiology* **18**, 52–58.
86. Gao S, Jin Y, Hall KS, *et al.* (2007) Selenium level and cognitive function in rural elderly Chinese. *Am J Epidemiol* **165**, 955–965.
87. Peretz A, Neve J, Desmedt J, Duchateau J, Dramaix M & Famaey JP (1991) Lymphocyte response is enhanced by supplementation of elderly subjects with selenium-enriched yeast. *Am J Clin Nutr* **53**, 1323–1328.
88. Kiremidjian-Schumacher L, Roy M, Wishe HI, Cohen MW & Stotzky G (1994) Supplementation with selenium and human immune cell functions. *Biol Trace Elem Res* **41**, 115–127.
89. Kiremidjian-Schumacher L, Roy M, Glickman R, Schneider K, Rothstein S, Cooper J, Hochster H, Kim M & Newman R (2000) Selenium and immunocompetence in patients with head and neck cancer. *Biol Trace Elem Res* **73**, 97–111.
90. Broome CS, McArdle F, Kyle JA, Andrews F, Lowe NM, Hart CA, Arthur JR & Jackson MJ (2004) An increase in selenium intake improves immune function and poliovirus handling in adults with marginal selenium status. *Am J Clin Nutr* **80**, 154–162.
91. Yu SY, Zhu YJ & Li WG (1997) Protective role of selenium against hepatitis B virus and primary liver cancer in Qidong. *Biol Trace Elem Res* **56**, 117–124.
92. Yu MW, Horng IS, Hsu KH, Chiang YC, Liaw YF & Chen CJ (1999) Plasma selenium levels and the risk of hepatocellular carcinoma among men with chronic hepatitis virus infection. *Am J Epidemiol* **150**, 367–374.
93. Li W, Zhu Y, Yan X, Zhang Q, Li X, Ni Z, Shen Z, Yao H & Zhu J (2000) The prevention of primary liver cancer by selenium in high risk populations. *Zhonghua Yu Fang Yi Xue Za Zhi* **34**, 336–338.
94. Bjelakovic G, Nikolova D, Simonetti RG & Gluud C (2004) Antioxidant supplements for prevention of gastrointestinal cancers: a systematic review and meta-analysis. *Lancet* **364**, 1219–1228.
95. Baum MK, Shor-Posner G, Lai S, Zhang G, Lai H, Fletcher MA, Sauberlich H & Page JB (1997) High risk of HIV-related mortality is associated with selenium deficiency. *J Acquir Immune Defic Syndr Hum Retrovirol* **15**, 370–374.
96. Campa A, Shor-Posner G, Indacochea F, Zhang G, Lai H, Asthana D, Scott GB & Baum MK (1999) Mortality risk in selenium-deficient HIV-positive children. *J Acquir Immune Defic Syndr Hum Retrovirol* **20**, 508–513.
97. Burbano X, Miguez-Burbano MJ, McCollister K, Zhang G, Rodriguez A, Ruiz P, Lecusay R & Shor-Posner G (2002) Impact of a selenium chemoprevention clinical trial on hospital admissions of HIV-infected participants. *HIV Clin Trials* **3**, 483–491.
98. Hurwitz BE, Klaus JR, Llabre MM, Gonzalez A, Lawrence PJ, Maher KJ, Greeson JM, Baum MK, Shor-Posner G, Skyler JS & Schneiderman N (2007) Suppression of human immunodeficiency virus type 1 viral load with selenium supplementation: a randomized controlled trial. *Arch Intern Med* **167**, 148–154.
99. Kupka R, Msamanga GI & Spiegelman D (2004) Se status is associated with accelerated HIV disease progression among HIV-infected pregnant women in Tanzania. *J Nutr* **134**, 2556–2560.
100. Scott R & MacPherson A (1998) Selenium supplementation in sub-fertile human males. *Br J Urol* **82**, 76–80.
101. Barrington JW, Lindsay P, James D, Smith S & Roberts A (1996) Selenium deficiency and miscarriage: a possible link? *Br J Obstet Gynaecol* **103**, 130–132.
102. Barrington JW, Taylor M, Smith S & Bowen-Simpkins P (1997) Selenium and recurrent miscarriage. *J Obstet Gynaecol* **17**, 199–200.
103. Rayman MP, Bode P & Redman CW (2003) Low selenium status is associated with the occurrence of the pregnancy disease preeclampsia in women from the United Kingdom. *Am J Obstet Gynecol* **189**, 1343–1349.
104. Zhuo H, Smith AH & Steinmaus C (2004) Selenium and lung cancer: a quantitative analysis of heterogeneity in the current

- epidemiological literature. *Cancer Epidemiol Biomarkers Prev* **13**, 771–778.
105. Zeegers MP, Goldbohm RA, Bode P & van den Brandt (2002) Prediagnostic toenail selenium and risk of bladder cancer. *Cancer Epidemiol Biomarkers Prev* **11**, 1292–1297.
 106. Brinkman M, Buntinx F, Muls E & Zeegers MP (2006) Use of selenium in chemoprevention of bladder cancer. *Lancet Oncol* **7**, 766–774.
 107. Wei WQ, Abnet CC, Qiao YL, Dawsey SM, Dong ZW, Sun XD, Fan JH, Gunter EW, Taylor PR & Mark SD (2004) Prospective study of serum selenium concentrations and esophageal and gastric cardia cancer, heart disease, stroke, and total death. *Am J Clin Nutr* **79**, 80–85.
 108. Rayman MP (2005) Selenium in cancer prevention: a review of the evidence and mechanism of action. *Proc Nutr Soc* **64**, 527–542.
 109. Etminan M, FitzGerald JM, Gleave M & Chambers K (2005) Intake of selenium in the prevention of prostate cancer: a systematic review and meta-analysis. *Cancer Causes Control* **16**, 1125–1131.
 110. Brinkman M, Reulen RC, Kellen E, Buntinx F & Zeegers MP (2006) Are men with low selenium levels at increased risk of prostate cancer? *Eur J Cancer* **42**, 2463–2471.
 111. Jacobs ET, Jiang R, Alberts DS, *et al.* (2004) Selenium and colorectal adenoma: results of a pooled analysis. *J Natl Cancer Inst* **96**, 1669–1675.
 112. Duffield-Lillico AJ, Reid ME, Turnbull BW, Combs GF Jr, Slate EH, Fischbach LA, Marshall JR & Clark LC (2002) Baseline characteristics and the effect of selenium supplementation on cancer incidence in a randomized clinical trial: a summary report of the Nutritional Prevention of Cancer Trial. *Cancer Epidemiol Biomarkers Prev* **11**, 630–639.
 113. Duffield-Lillico AJ, Dalkin BL, Reid ME, Turnbull BW, Slate EH, Jacobs ET, Marshall JR, Clark LC & Nutritional Prevention of Cancer Study Group (2003) Selenium supplementation, baseline plasma selenium status and incidence of prostate cancer: an analysis of the complete treatment period of the Nutritional Prevention of Cancer Trial. *BJU Int* **91**, 608–612.
 114. Reid ME, Duffield-Lillico AJ, Garland L, Turnbull BW, Clark LC & Marshall JR (2002) Selenium supplementation and lung cancer incidence: an update of the Nutritional Prevention of Cancer trial. *Cancer Epidemiol Biomarkers Prev* **11**, 1285–1291.
 115. Reid ME, Duffield-Lillico AJ, Sunga A, Fakh M, Alberts DS & Marshall JR (2006) Selenium supplementation and colorectal adenomas: an analysis of the Nutritional Prevention of Cancer trial. *Int J Cancer* **118**, 1777–1781.
 116. Gärtner R, Gasier BC, Dietrich JW, Krebs B & Angsturm MW (2002) Selenium supplementation in patients with autoimmune thyroiditis decreases thyroid peroxidase antibodies concentrations. *J Clin Endocrinol Metab* **87**, 1687–1691.
 117. Duntas LH, Mantzou E & Koutras DA (2003) Effects of six month treatment with selenomethionine in patients with autoimmune thyroiditis. *Eur J Endocrinol* **148**, 389–393.
 118. Turker O, Kumanlioglu K, Karapolat I & Dogan I (2006) Selenium treatment in autoimmune thyroiditis: 9-month follow-up with variable doses. *J Endocrinol* **190**, 151–156.
 119. Derumeaux H, Valeix P, Castetbon K, Bensimon M, Boutron-Ruault MC, Arnaud J & Hercberg S (2003) Association of selenium with thyroid volume and echostucture in 35- to 60-year-old French adults. *Eur J Endocrinol* **148**, 309–315.
 120. Glattre E, Thomassen Y, Thoresen SO, Haldorsen T, Lund-Larsen PG, Theodorsen L & Aaseth J (1989) Prediagnostic serum selenium in a case-control study of thyroid cancer. *Int J Epidemiol* **18**, 45–49.
 121. Jellum E, Andersen A, Lund-Larsen P, Theodorsen L & Orjasaeter H (1993) The JANUS serum bank. *Sci Total Environ* **139–140**, 527–535.
 122. Negro R, Greco G, Mangieri T, Pezzarossa A, Dazzi D & Hassan H (2007) The influence of selenium supplementation on postpartum thyroid status in pregnant women with thyroid peroxidase autoantibodies. *J Clin Endocrinol Metab* **92**, 1263–1268.
 123. Flores-Mateo G, Navas-Acien A, Pastor-Barriuso R & Guallar E (2006) Selenium and coronary heart disease: a meta-analysis. *Am J Clin Nutr* **84**, 762–773.
 124. Arteel GE, Briviba K & Sies H (1999) Protection against peroxynitrite. *FEBS Lett* **445**, 226–230.
 125. Xia Y, Hill KE, Byrne DW, Xu J & Burk RF (2005) Effectiveness of selenium supplements in a low-selenium area of China. *Am J Clin Nutr* **81**, 829–834.
 126. Burk RF, Hill KE, Motley AK, Austin LM & Norworthy BK (2006) Deletion of selenoprotein P upregulates urinary selenium excretion and depresses whole-body selenium content. *Biochim Biophys Acta* **1760**, 1789–1793.
 127. Hill KE, Zhou J, McMahan WJ, Motley AK & Burk RF (2004) Neurological dysfunction occurs in mice with targeted deletion of the selenoprotein P gene. *J Nutr* **134**, 157–161.
 128. Valentine WM, Hill KE, Austin LM, Valentine HL, Goldowitz D & Burk RF (2005) Brainstem axonal degeneration in mice with deletion of selenoprotein P. *Toxicol Pathol* **33**, 570–576.
 129. Dong Y, Lisk D, Block E & Ip C (2001) Characterization of the biological activity of γ -glutamyl-Se-methylselenocysteine: a novel, naturally occurring anticancer agent from garlic. *Cancer Res* **61**, 2923–2928.
 130. Ip C (1998) Lessons from basic research in selenium and cancer prevention. *J Nutr* **128**, 1845–1854.
 131. Zeng H & Combs GF Jr (2008) Selenium as an anticancer nutrient: roles in cell proliferation and tumor cell invasion. *J Nutr Biochem* **19**, 1–7.
 132. Finley JW (2003) Reduction of cancer risk by consumption of selenium-enriched plants: enrichment of broccoli with selenium increases the anticarcinogenic properties of broccoli. *J Med Food* **6**, 19–26.
 133. Bates CJ, Thane CW, Prentice A & Delves HT (2002) Selenium status and its correlates in a British National Diet and Nutrition Survey: people aged 65 years and over. *J Trace Elem Med Biol* **6**, 1–8.
 134. Hesse-Bahr K, Dreher I & Kehrle J (2000) The influence of the cytokines IL-1 β and INF γ on the expression of selenoproteins in the human hepatocarcinoma cell line HepG2. *Biofactors* **11**, 83–85.
 135. Bjelakovic G, Nikolova D, Glud LL, Simonetti RG & Glud C (2007) Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. *JAMA* **297**, 842–857.
 136. Papp LV, Lu J, Holmgren A & Khanna KK (2007) From selenium to selenoproteins: synthesis, identity, and their role in human health. *Antioxid Redox Signal* **9**, 775–806.
 137. Rayman MP (2000) The importance of selenium to human health. *Lancet* **356**, 233–241.
 138. Schweizer U, Bräuer AU, Köhrle J, Nitsch R & Savaskan NE (2004) Selenium and brain function: a poorly recognized liaison. *Brain Res Brain Res Rev* **45**, 164–178.
 139. Schiavon R, Guidi GC, Biasioli S, De Fanti E & Targa L (1994) Plasma glutathione peroxidase activity as an index of renal function. *Eur J Clin Chem Clin Biochem* **32**, 759–765.
 140. Ravaglia G, Forti P, Maioli F, Muscarelli A, Sacchetti L, Arnone G, Nativio V, Talerico T & Mariani E (2003) Homocysteine and cognitive function in healthy elderly community dwellers in Italy. *Am J Clin Nutr* **77**, 668–673.
 141. Gori AM, Corsi AM, Fedi S, Gazzini A, *et al.* (2005) A proinflammatory state is associated with hyperhomocysteinemia in the elderly. *Am J Clin Nutr* **82**, 335–341.

142. Beck MA, Esworthy RS, Ho Y-S & Chu F-F (1998) Glutathione peroxidase protects mice from viral-induced myocarditis. *FASEB J* **12**, 1143–1149.
143. Zhao L, Cox AG, Ruzicka JA, Bhat AA, Zhang W & Taylor EW (2000) Molecular modeling and in vitro activity of an HIV-1-encoded glutathione peroxidase. *Proc Natl Acad Sci U S A* **97**, 6356–6361.
144. Zhang W, Ramanathan CS, Nadimpalli RG, Bhat AA, Cox AG & Taylor EW (1999) Selenium-dependent glutathione peroxidase modules encoded by RNA viruses. *Biol Trace Elem Res* **70**, 97–116.
145. Shisler JL, Senkevich TG, Berry MJ & Moss B (1998) Ultraviolet-induced cell death blocked by a selenoprotein from a human dermatotropic poxvirus. *Science* **279**, 102–105.
146. Maiorino M & Ursini F (2002) Oxidative stress, spermatogenesis and fertility. *Biol Chem* **383**, 591–597.
147. Pfeifer H, Conrad M, Roethlein D, Kyriakopoulos A, Brielmeyer M, Bornkamm G & Behne D (2001) Identification of a specific sperm nuclei selenoenzyme necessary for protamine thiol cross-linking during sperm maturation. *FASEB J* **15**, 1236–1238.
148. Flohé L (2007) Selenium in mammalian spermiogenesis. *Biol Chem* **388**, 987–995.
149. Behne D, Kyriakopoulos A, Kalcklösch M, Weiss-Nowak C, Pfeifer H, Gessner H & Hammel C (1997) Two new selenoproteins found in the prostatic glandular epithelium and in the spermatid nuclei. *Biomed Environ Sci* **10**, 340–345.
150. Brigelius-Flohé R (2006) Glutathione peroxidases and redox-regulated transcription factors. *Biol Chem* **387**, 1329–1335.
151. Hawkes WC & Turek PJ (2001) Effects of dietary selenium on sperm motility in healthy men. *J Androl* **22**, 764–772.
152. Nomura AM, Lee J, Stemmermann GN & Combs GF Jr (2000) Serum selenium and subsequent risk of prostate cancer. *Cancer Epidemiol Biomarkers Prev* **9**, 883–887.
153. Levander OA, Alfthan G, Arvilommi H, Gref CG, Huttunen JK, Kataja M, Koivistoinen P & Pikkariainen J (1983) Bioavailability of selenium to Finnish men as assessed by platelet glutathione peroxidase activity and other blood parameters. *Am J Clin Nutr* **37**, 887–897.
154. Rayman M, Thompson A, Warren-Perry M, Galassini R, Catterick J, Hall E, Lawrence D & Bliss J (2006) Impact of selenium on mood and quality of life: a randomized, controlled trial. *Biol Psychiatry* **59**, 147–154.
155. Duffield-Lillico AJ, Slate EH, Reid ME, *et al.* (2003) Nutritional Prevention of Cancer Study Group. Selenium supplementation and secondary prevention of nonmelanoma skin cancer in a randomized trial. *J Natl Cancer Inst* **95**, 1477–1481.
156. Stranges S, Marshall JR, Natarajan R, Donahue RP, Trevisan M, Combs GF, Cappuccio FP, Ceriello A & Reid ME (2007) Effects of long-term selenium supplementation on the incidence of type 2 diabetes: a randomized trial. *Ann Intern Med* **147**, 217–223.
157. Burk RF, Norworthy BK, Hill KE, Motley AK & Byrne DW (2006) Effects of chemical form of selenium on plasma biomarkers in a high-dose human supplementation trial. *Cancer Epidemiol Biomarkers Prev* **15**, 804–810.
158. Peters U, Foster CB, Chatterjee N, Schatzkin A, Reding D, Andriole GL, Crawford ED, Sturup S, Chanock SJ & Hayes RB (2007) Serum selenium and risk of prostate cancer – a nested case–control study. *Am J Clin Nutr* **85**, 209–217.
159. Beckett GJ & Arthur JR (2005) Selenium and endocrine systems. *J Endocrinol* **184**, 455–465.
160. Korpela H, Kumpulainen J, Jussila E, Kemilä S, Kääriäinen M, Kääriäinen T & Sotaniemi EA (1989) Effect of selenium supplementation after acute myocardial infarction. *Res Commun Chem Pathol Pharmacol* **65**, 249–252.
161. Stranges S, Marshall JR, Trevisan M, Natarajan R, Donahue RP, Combs GF, Farinero E, Clark LC & Reid ME (2006) Effects of selenium supplementation on cardiovascular disease incidence and mortality: secondary analyses in a randomized clinical trial. *Am J Epidemiol* **163**, 694–699.
162. Brooks JD, Metter EJ, Chan DW, Sokoll LJ, Landis P, Nelson WG, Muller D, Andres R & Carter HB (2001) Plasma selenium level before diagnosis and the risk of prostate cancer development. *J Urol* **166**, 2034–2038.
163. Li B, Taylor PR, Li JY, *et al.* (1993) Linxian nutrition intervention trials. Design, methods, participant characteristics, and compliance. *Ann Epidemiol* **3**, 577–585.
164. Vanderpas JB, Contempré B, Duale NL, Deckx H, Bebe N, Longombé AO, Thilly CH, Diplock AT & Dumont JE (1993) Selenium deficiency mitigates hypothyroxinemia in iodine-deficient subjects. *Am J Clin Nutr* **57**, Suppl. 2, 271S–275S.
165. Burk RF, Early DS, Hill KE, Palmer IS & Boeglin ME (1998) Plasma selenium in patients with cirrhosis. *Hepatology* **27**, 794–798.
166. Mostert V, Dreher I, Kohrle J & Abel J (1999) Transforming growth factor- β 1 inhibits expression of selenoprotein P in cultured human liver cells. *FEBS Lett* **460**, 23–26.
167. Dreher I, Jakobs TC & Kohrle J (1997) Cloning and characterization of the human selenoprotein P promoter. Response of selenoprotein P expression to cytokines in liver cells. *J Biol Chem* **272**, 29364–29371.
168. Walter PL, Steinbrenner H, Barthel A & Klotz LO (2008) Stimulation of selenoprotein P promoter activity in hepatoma cells by FoxO1a transcription factor. *Biochem Biophys Res Commun* **365**, 316–321.
169. Irons R, Carlson BA, Hatfield DL & Davis CD (2006) Both selenoproteins and low molecular weight seleno compounds reduce colon cancer risk in mice with genetically impaired selenoprotein expression. *J Nutr* **136**, 1311–1317.
170. Brown KM, Pickard K, Nicol F, Beckett GJ, Duthie GG & Arthur JR (2000) Effects of organic and inorganic selenium supplementation on selenoenzyme activity in blood lymphocytes, granulocytes, platelets and erythrocytes. *Clin Sci (Lond)* **98**, 593–599.
171. Hu YJ & Diamond AM (2003) Role of glutathione peroxidase 1 in breast cancer: loss of heterozygosity and allelic differences in the response to selenium. *Cancer Res* **63**, 3347–3351.
172. Kumaraswamy E, Malykh A, Korotkov KV, Kozyavkin S, Hu Y, Kwon SY, Moustafa ME, Carlson BA, Berry MJ, Lee BJ, Hatfield DL, Diamond AM & Gladyshev VN (2000) Structure–expression relationships of the 15-kDa selenoprotein gene. Possible role of the protein in cancer etiology. *J Biol Chem* **275**, 35540–35547.
173. Ratnasinghe D, Tangrea JA, Andersen MR, Barrett MJ, Virtamo J, Taylor PR & Albanes D (2000) Glutathione peroxidase codon 198 polymorphism variant increases lung cancer risk. *Cancer Res* **60**, 6381–6383.
174. Méplan C, Crosley LK, Nicol F, Beckett GJ, Howie AF, Hill KE, Horgan G, Mathers JC, Arthur JR & Hesketh JE (2007) Genetic polymorphisms in the human selenoprotein P gene determine the response of selenoprotein markers to selenium supplementation in a gender-specific manner (the SELGEN study). *FASEB J* **21**, 3063–3074.
175. Ichimura Y, Habuchi T, Tsuchiya N, Wang L, Oyama C, Sato K, Nishiyama H, Ogawa O & Kato T (2004) Increased risk of bladder cancer associated with a glutathione peroxidase 1 codon 198 variant. *J Urol* **172**, 728–732.
176. Ravn-Haren G, Olsen A, Tjønneland A, Dragsted LO, Nexø BA, Wallin H, Overvad K, Raaschou-Nielsen O & Vogel U (2006) Associations between GPX1 Pro198Leu polymorphism, erythrocyte GPX activity, alcohol consumption and breast

- cancer risk in a prospective cohort study. *Carcinogenesis* **27**, 820–825.
177. Lee OJ, Schneider-Stock R, McChesney PA, Kuester D, Roessner A, Vieth M, Moskaluk CA & El-Rifai W (2005) Hypermethylation and loss of expression of glutathione peroxidase-3 in Barrett's tumorigenesis. *Neoplasia* **7**, 854–861.
 178. Chen Y, Hall M, Graziano JH, Slavkovich V, van Geen A, Parvez F & Ahsan H (2007) A prospective study of blood selenium levels and the risk of arsenic-related premalignant skin lesions. *Cancer Epidemiol Biomarkers Prev* **16**, 207–213.
 179. Arnaud J, Akbaraly NT, Hininger I, Roussel AM & Berr C (2007) Factors associated with longitudinal plasma selenium decline in the elderly: the EVA study. *J Nutr Biochem* **18**, 482–487.
 180. Euroala M & Hietaniemi V (2000) *Report of the Selenium Monitoring Programme 1997–1999. Publications of Agricultural Research Centre of Finland, Series B 24*. Jokioinen: Agricultural Research Centre of Finland.
 181. Hartikainen H (2005) Biogeochemistry of selenium and its impact on food chain quality and human health. *J Trace Elem Med Biol* **18**, 309–318.
 182. Aro A, Alfthan G, Ekholm P & Varo P (1998) Effects of selenium supplementation of fertilizers on human nutrition and selenium status. In *Environmental Chemistry of Selenium*, pp. 81–97 [WT Frankenberger Jr and RA Engberg, editors]. New York: Marcel Dekker, Inc.
 183. Rayman MP, Angus F & Goenaga Infante H (2007) Bioavailability and speciation of selenium from selenium-enriched mushrooms. *Proc Nutr Soc* **66**, 55A.
 184. Cole DJA (2000) Selenium, the pig and the human diet. In *Concepts in Pig Science*, pp. 149–158 [TP Lyons and DJA Cole, editors]. Nottingham, UK: Nottingham University Press.
 185. Lyons MP, Papazyan TT & Surai PF (2007) Selenium in food chain and animal nutrition: lessons from nature. *Asian-Aust J Anim Sci* **20**, 1135–1155.
 186. Arnold WN & Thrasher JB (2003) Selenium concentration in the prostate. *Biol Trace Elem Res* **91**, 277–280.
 187. Gianduzzo TR, Holmes EG, Tinggi U, Shahin M, Mactaggart P & Nicol D (2003) Prostatic and peripheral blood selenium levels after oral supplementation. *J Urol* **170**, 870–873.
 188. Sabichi AL, Lee JJ, Taylor RJ, *et al.* (2006) Selenium accumulation in prostate tissue during a randomized, controlled short-term trial of L-selenomethionine: a Southwest Oncology Group Study. *Clin Cancer Res* **12**, 2178–2184.
 189. Taylor PR & Greenwald P (2005) Nutritional interventions in cancer prevention. *J Clin Oncol* **23**, 333–345.
 190. Niskar AS, Paschal DC, Kieszak SM, *et al.* (2003) Serum selenium levels in the US population: Third National Health and Nutrition Examination Survey, 1988–1994. *Biol Trace Elem Res* **91**, 1–10.
 191. Lawson KA, Wright ME, Subar A, Mouw T, Hollenbeck A, Schatzkin A & Leitzmann MF (2007) Multivitamin use and risk of prostate cancer in the National Institutes of Health-AARP Diet and Health Study. *J Natl Cancer Inst* **99**, 754–764.